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McAN: a novel computational algorithm and platform for constructing and visualizing haplotype networks

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Abstract

Haplotype networks are graphs used to represent evolutionary relationships between a set of taxa and are characterized by intuitiveness in analyzing genealogical relationships of closely related genomes. We here propose a novel algorithm termed McAN that considers mutation spectrum history (mutations in ancestry haplotype should be contained in descendant haplotype), node size (corresponding to sample count for a given node) and sampling time when constructing haplotype network. We show that McAN is two orders of magnitude faster than state-of-the-art algorithms without losing accuracy, making it suitable for analysis of a large number of sequences. Based on our algorithm, we developed an online web server and offline tool for haplotype network construction, community lineage determination, and interactive network visualization. We demonstrate that McAN is highly suitable for analyzing and visualizing massive genomic data and is helpful to enhance the understanding of genome evolution. Availability: Source code is written in C/C++ and available at https://github.com/Theory-Lun/McAN and https://ngdc.cncb.ac.cn/biocode/tools/BT007301 under the MIT license. Web server is available at https://ngdc.cncb.ac.cn/bit/hapnet/. SARS-COV-2 dataset are available at https://ngdc.cncb.ac.cn/ncov/. Contact: songshh@big.ac.cn (Song S), zhaowm@big.ac.cn (Zhao W), baoym@big.ac.cn (Bao Y), zhangzhang@big.ac.cn (Zhang Z), ybxue@big.ac.cn (Xue Y).

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INTRODUCTION

Understanding how species evolve over millions of years requires the processing of massive amounts of genomic data, which would be a challenge without powerful bioinformatic tools. One of these tools, haplotype networks, plays important roles in tracing the evolution and migration of diverse species and is fundamental to determine and visualize genealogical relationships of population genomes [1, 2]. Over the past decades, several algorithms have been proposed for haplotype network construction, including

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This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (https://creativecommons.org/ licenses/by-nc/4.0/), which permits non-commercial re-use, distribution, and reproduction in any medium, provided the original work is properly cited. For commercial re-use, please contact journals.permissions@oup.com minimum spanning network (MSN) [3], median-joining network (MJN) [3], Templeton Crandall and Sing algorithm (TCS) [4] and randomized minimum spanning tree (RMST) [5]. These algorithms have been applied to analyze a variety of genome sequences, including virus genomes, human mitochondria [1], plant chloroplast [2] and mammalian Y chromosome [6]. Haplotype network is extensively used in viral studies, with the aim to identify possible transmission routes of viruses, including the virus responsible for the ongoing COVID-19 pandemic [7–11].

Typically, these algorithms use a haplotype distance matrix to construct network, which are crucial for inferring ancestrydescendant relationships between haplotypes; however, they are limited in many ways. First, they ignore several important features when constructing haplotype network, such as sampling time, number of sequences in haplotypes and mutation spectrum history, which can mislead the tracing of evolution/transmission routes. Second, they generate undirected networks and thus are unable to reflect the evolutionary directions of variants. Third, running these algorithms is time-consuming and memoryintensive. The time complexities of MSN, MJN and TCS are $O(m^2)$ (worst case), $O(n^{2.2})$ (average case) and $O(m^5)$ (worst case), respectively, where *m* is the number of haplotypes and *n* is the number of sequences (Supplementary Table S1). Therefore, these algorithms are incapable of processing the massive datasets, such as the tens of millions of SARS-CoV-2 sequences currently available. In practice, MJN takes more than three days to build haplotype networks for 1000 SARS-CoV-2 sequences, mainly due to the calculation of distance between any two haplotypes.

The visualization of haplotype networks is also vital for tracing and analysis of the genealogical relationships. In recent decades, various frameworks for network visualization have emerged. For instance, igraph [12], Gephi [13], Pajek [14] and Tulip [15] provide network visualization services for desktop users, and WiGis [16] for large graphs online. Furthermore, visualization platforms specific for multi-dimensional biological data have also been developed, such as Cytoscape [17], VisANT [18], PopArt [19], CoV Genome Tracker [20] and TPD [21]. However, most of them are not capable of fully supporting large-scaled genome data or dynamically fine-tuning network layouts.

To address these issues, we proposed a new haplotype network construction algorithm called Minimum-cost Arborescence Network (McAN), which is designed to build a directed, rooted tree spanning all vertices with minimum cost by taking into account both genome-wide mutation spectrum features and epidemiological characteristics, and is capable to run two orders of magnitude faster than existing algorithms. Moreover, we developed a web-based haplotype network visualization platform, which integrates spatial-temporal information and supports determination of community lineage and interactive fine-tuning of network layouts. We evaluated the performance of our algorithm and platform on both simulated datasets and multiple real datasets, which illustrated the advantage of McAN in analyzing and visualizing massive genomic data.

MATERIALS AND METHODS Overview of the method

McAN was built based on minimum-cost arborescence for haplotype network construction, a well-known graph theory widely used in multiple applications. In McAN, haplotype network is represented by a directed, rooted arborescence [22], in which each haplotype is a cluster of identical sequences (with same mutations after filtering), each edge reflects directed ancestordescendant relationship between haplotypes, and optimal arborescence is determined by minimum distances of all summed edges.

McAN factors in three features when constructing a haplotype network, including mutation spectrum histories, sample sizes and sampling times for every haplotype. Specifically, McAN takes account of the mutation spectrum history of all samples and assumes that mutations of an ancestral haplotype should be inherited by its descendant haplotypes, and haplotypes with more sequences should be more likely to be intermediate ancestor nodes, due to a higher probability of propagating outwards of the haplotypes with more sequences. If sampling time is available, ancestral mutants are assumed to be earlier sampled than derived mutants.

For convenience, these assumptions are summarized as the following four criteria (Figure 1): (1) mutation spectrum history, (2) large ancestry haplotype, (3) ancestry earlier sampling time and (4) minimum evolution (ME). The last criterion means the optimal haplotype network for a set of haplotypes is assumed to be an arborescence whose sum of distances of all directed edges is minimal [23]. We defined the distance between a pair of haplotypes as the number of different mutations between two haplotypes.

When all data including mutation and metadata are read by McAN, pairs of accession ID and mutations for all sequences are stored in a hash map, and sequences with the same mutations are clustered into a group and regarded as a haplotype. The reference sequence (users need to select the earliest high-quality sequence as reference) is designated as the root node of the haplotype network. Next, all haplotypes are sorted by mutation count and sequence count in descending order and the earliest sampling time (if available) in ascending order (Supplementary Figure S1A). Based on this sorting, the closest ancestor is determined and minimized for each haplotype (Supplementary Figure S1B). To save memory and running time, McAN calculates distances between adjacent haplotypes instead of any two haplotypes (Supplementary Figure S1C). Then, the directed edges representing the candidate immediate ancestor-descendant relationships are established under the constraint of the 'mutation spectrum history' criterion. The immediate ancestor-descendant relationships are determined by the 'minimum evolution (ME)' criterion. In practice, the haplotype networks may not be unique, so we propose the 'large ancestry haplotype' and 'ancestry earlier sampling time' criterion to determine, which one is the best among all haplotype networks. In addition, we further paralleled the algorithm of McAN to satisfy huge amount sequences data.

Detailed algorithm of McAN

Suppose there are m haplotypes, denoted by $h_0,h_1,\ldots,h_{m-1},$ respectively. Let

$$V = \{h_i | i = 0, 1, \dots, m - 1\}$$
(1)

be the set of all haplotypes and $M_i = \{mut | mut = (pos, ref, alt)\}\$ be the set of mutations in haplotype h_i including all SNPs, insertions and deletions, where $pos \in \mathbb{Z}_+$ represents the genomic position of mutation mut and ref (alt) a reference (mutated) nucleotide base or a sub-sequence of the reference (mutated) sequence. Let h_r be the haplotype containing the reference sample. Obviously, the set of mutations M_r in haplotype h_r is equal to an empty set. According to the 'mutation spectrum history' criterion, the set of candidates of directed edges should then be

$$E = \{e_{ij} = (h_i, h_j) | h_i, h_j \in V, M_i \subsetneq M_j\},$$
(2)

where e_{ij} is a directed edge from haplotype h_i to h_j , which also be denoted as an ordered pair (h_i, h_j) or (i, j). Notice that, no



Figure 1. Schematic illustration for haplotype network construction by McAN. To infer the genealogical relationship among multiple genomic sequences, four criteria were adopted for haplotype network construction. Then, optimization problem was abstracted and the McAN algorithm was designed. Furthermore, McAN tool and online webserver was developed to implement this algorithm. Finally, a haplotype network was constructed and visualized interactively by McAN platform.

self-cycle belongs to set E. The distance between two haplotypes h_i and h_j satisfying $(h_i,h_j)\in E$ is

$$d_{ij} = |M_j| - |M_i|, (3)$$

where M_i and M_j are sets of mutations of haplotypes h_i and h_j , respectively, $|\cdot|$ denotes the cardinality of a set. Let $w_1(\cdot)$ be a cost function (or weight function) on E,

$$\begin{split} w_1 &: E \to \mathbb{Z}_+, \\ e_{ij} &\mapsto d_{ij}. \end{split}$$

By 'minimum evolution (ME)' criterion, the haplotype network must be the minimum-cost arborescence of the weighted directed graph $G_1 = (V, E, w_1)$. Formally, the minimum-cost arborescence problem can be represented as an integer programming problem:

$$\min_{\substack{x_{ij} \\ i,j \in E}} \sum_{\substack{(i,j) \in E \\ i \\ i \\ j}} d_{ij} x_{ij},$$
s.t. $\sum_{i} x_{ij} = 1, \forall j \in V \setminus r,$

$$\sum_{i,j} x_{ij} = m - 1,$$

$$x_{ij} \in \{0, 1\},$$
(5)

where x_{ij} , $(i,j) \in E$, are the decision variables, r is the index of haplotype containing reference. If edge (i,j) is in the minimum-cost arborescence, $x_{ij} = 1$, otherwise $x_{ij} = 0$. The first constraint

represents that the number of ancestors of haplotypes except for r should be 1, and the second one represents that the total number of edges in haplotype network must be m - 1. Note that a decision variable $x_{ij} \in \{0, 1\}$ satisfies these two constraints if and only if $\{e_{ij}|x_{ij} = 1, e_{ij} \in E\}$ is a spanning arborescence of the unweighted directed graph G = (V, E).

To deal with the minimum-cost arborescence problem (problem (5)), one should determine the root of arborescence, handle directed cycles in graph and examine the uniqueness of the solution. The root of minimum-cost arborescence is supposed to be provide by the use, which is the haplotype h_r containing the reference sequence. The following Theorem 1 shows that there is no directed cycle in G_1 .

Theorem 1. Let G = (V, E) be a directed graph, where V and E is defined by Equations (1) and (2), respectively. Then, there is no directed cycle in G.

Proof. Suppose for contradiction that there exists at least one directed cycle in *G*. Then, let $C = v_0 a_1 v_1 \cdots a_k v_k$ be a directed cycle in *G*, where $v_0, v_1, \cdots, v_k \in V$, $a_1, \cdots, a_k \in E$, $a_i = (v_{i-1}, v_i)$ and $v_0 = v_k$. And let \tilde{M}_i be the set of mutations in vertex (or haplotype) v_i , for any $i = 0, 1, \cdots, k$. By Equation (2), we have $\tilde{M}_{i-1} \subsetneq \tilde{M}_i$, $i = 1, 2, \cdots, k$. Therefore, we obtain $\tilde{M}_0 \subsetneq \tilde{M}_k$. However, \tilde{M}_0 is equal to \tilde{M}_k since v_0 is equal to v_k . This is a contradiction. Hence, our assumption that "there exists at least one directed cycle in *G*" cannot be true. We thus have proved "there is no directed cycle in *G*". Q.E.D.

However, the minimum-cost arborescence of G_1 , may not be unique. Therefore, two other criteria are involved in to further constrain possible solutions. If more than one haplotype exists that is both closest ancestry of a given haplotype, then the haplotype containing more sequences is chosen to be the ancestry of the given haplotype by 'large ancestry haplotype' criterion. If more than one haplotype exists that is both closest ancestry of a given haplotype and is both meeting 'large ancestry haplotype' criterion, then the one meeting the 'ancestry earlier sampling time' criterion is retained in the haplotype network. Formally, let $w_2(\cdot)$ be a cost function on E

$$\begin{aligned} w_2 &: E \to \mathbb{Z}^2_+ \times \{0, 1\}, \\ e_{ij} &\mapsto (d_{ij}, z_i, \tilde{t}_{ij}), \end{aligned}$$
 (6)

where z_i is the number of sequences in h_i , \tilde{t}_{ij} , $(i, j) \in E$, is defined as

$$\tilde{t}_{ij} = \begin{cases} 1, t_i \le t_j, \\ 0, t_i > t_j, \end{cases}$$
(7)

representing whether edge (h_i, h_j) in *E* meet 'ancestry earlier sampling time' criterion, t_i is the sampling time of the ith haplotype h_i (which is defined as the earliest sampling time of sequences in h_i). Considering all the four criteria, the haplotype network is a minimum spanning arborescence of $G_2 = (V, E, w_2)$ with order relation \prec , where order relation \prec is defined on $\mathbb{Z}^2_+ \times \{0, 1\}$ such that for any $a = (d_1, z_1, \tilde{t}_1)$ and $b = (d_2, z_2, \tilde{t}_2)$ in $\mathbb{Z}^2_+ \times \{0, 1\}, a \prec b$ if and only if $d_1 < d_2$ or $(d_1 = d_2$ and $z_1 > z_2)$ or $(d_1 = d_2$ and $z_1 = z_2$ and $\tilde{t}_1 \ge \tilde{t}_2$). The overall optimization problem becomes

$$\min_{x_{ij}} \sum_{(i,j) \in E} w_2(e_{ij}) x_{ij},$$

s.t. $\sum_{i} x_{ij} = 1, \forall j \in V \setminus r,$
 $\sum_{i,j} x_{ij} = m - 1,$
 $x_{ij} \in \{0, 1\},$ (8)

where x_{ij} , $(i, j) \in E$, are the decision variables.

Optimization problem (8) is not a standard minimum-cost arborescence problem with a numerical cost, but with a vectorial cost. However, we can follow the framework of Chu-Liu-Edmonds' algorithm [24, 25] to solve problem (8), although we cannot use the algorithm directly. Specifically, we firstly find all the edges in *E*, and calculate the distance for all edges in *E*. Next, we find an edge (h_j, h_i) , incoming to h_i with the lowest cost (according to relation \prec , and weight $w_2(\cdot)$) for each haplotype $h_i \in V \setminus \{h_r\}$, where h_r represent the haplotype containing the reference, $j = \pi(i)$ is defined as

$$\pi(\mathbf{i}) = \arg \min_{\mathbf{j}} w_2(e_{\mathbf{j}\mathbf{i}}). \tag{9}$$

The pseudo-code of constructing haplotype network by following the framework of Chu-Liu-Edmonds' algorithm is represented as Algorithm S1. The output A generated by Algorithm S1 must be an optimum arborescence of problem (8). Chu-Liu-Edmonds' algorithm takes the set of edges E and the weight of each edge as input. However, the input of constructing haplotype network are the mutations set, the sampling time and the number of samples of each haplotype. If we directly use the framework of Chu-Liu-Edmonds' algorithm to solve the problem of constructing haplotype network, the computation of determining if $M_i \subsetneq M_j$ for each pair of (i, j) satisfying $|M_i| < |M_j|$ cannot be saved. Meanwhile

the storage of the weight $w_2(e_{ij})$ (that is d_{ij} and \tilde{t}_{ij} in Algorithm S1) of each edge in *E* cannot be reduced. Therefore, both best-case and worst-case time complexity of Algorithm S1 are $O(|V|^2)$ and space complexity of Algorithm S1 are O(|E|)

To reduce the time and space complexity of Algorithm S1, we proposed McAN (Algorithm 1). McAN firstly sort all haplotypes by the number of mutations and sequences in descending order, and the earliest sampling time in ascending order. Haplotypes and their mutation sets under the new order are denoted by $\{h_{s(i)}\}$ and $\{M_{s(i)}\}$, i = 0, 1, ..., m - 1, respectively, where s(i) is a permutation of set $\{0, 1, ..., m - 1\}$. For each haplotype $h_{s(i)}$, its directed ancestor $h_{s(j)}$ is determined by searching from j = i + 1 and finding the smallest j satisfying $M_{s(j)} \subsetneq M_{s(j)}$, that is

 $\pi^{*}(i) = \min\{j | M_{s(j)} \subsetneq M_{s(i)}, i+1 \le j \le m-1\}, i = 0, 1, \dots, m-2.$ (10)

The pseudo-code of the proposed McAN is represented as Algorithm 1, where sort is a sorting algorithms (sort generate the permutation s(i) remaining $\{M_i\}, \{z_i\}$ and $\{t_i\}$ unchanged), and \prec is an order relation defined on $\mathbb{N} \times \mathbb{Z}_+^2$ such that for any $a = (|M_1|, z_1, t_1)$ and $b = (|M_2|, z_2, t_2), a \prec b$ if and only if $|M_1| > |M_2|$ or $(|M_1| = |M_2|$ and $z_1 > z_2)$ or $(|M_2| = |M_2|$ and $z_1 = z_2$ and $t_1 \leq t_2$). So, we have

$$(M_{s(k)}, Z_{s(k)}, t_{s(k)}) \prec (M_{s(l)}, Z_{s(l)}, t_{s(l)}), 0 \le k \le l \le m - 1.$$
(11)

The output A of Algorithm 1 is proved to be an optimal solution of problem (8) by Theorem 2 (Lemma 1–3 in the proof of Theorem 2 is provided in the supplementary materials).

Algorithm 1. McAN

Input:
m, the number of haplotypes,
M_i , $i = 0, 1, \dots, m - 1$, the set of mutations in haplotype h_i ,
t_i , $i = 0, 1, \dots, m - 1$, the sampling time of haplotype h_i ,
z_i , $i = 0, 1,, m - 1$, the number of samples in haplotype h_i .
Output:
A, the minimum-cost arborescence, which is a set of edges.
Step 1 (sort all haplotypes by generating a permutation {s(k)})
$\{s(k)\} \leftarrow sort(\{(M_i , z_i, t_i)\}, \prec).$
Step 2 (find minimum-cost arborescence A)
$A \leftarrow \emptyset$,
for each i from 0 to $m - 2$,
for each j from $i + 1$ to $m - 1$,
if $ M_{s(i)} \neq M_{s(i)} $, then
if $M_{s(i)} \subseteq M_{s(i)}$, then
$A \leftarrow A \cup \{(s(j), s(i))\},\$
break,
end if, end if, end for, end for,
return A.

Theorem 2. The output A[⋆] of Algorithm 1 is an optimal solution of problem (8).

Proof. First, we will show that A^* is a feasible solution of problem (8). For each $i = 0, 1, \ldots, m - 2$, by Lemma 1, we have $M_{s(m-1)} = \emptyset \subsetneq M_{s(i)}$. Therefore, for any $i = 0, 1, \ldots, m - 2$ in step 2 of Algorithm 1, there exists at least one *j*, satisfying j > i and $M_{s(j)} \subsetneq M_{s(i)}.$ Then the output of Algorithm 1 can be represented as

$$A^{\star} = \{ (s(\pi^{\star}(i)), s(i)) | i = 0, 1, \dots, m-2 \},$$
(12)

where $\pi^{\star}(i)$ is defined by Equation (10). Then we have

$$\sum_{(k,l)\in E} x_{kl}^{\star} = |A^{\star}| = m-1,$$

and

$$\sum_{k} x_{kl}^{\star} = \begin{cases} 1, l \in V \setminus r, \\ 0, l = r, \end{cases}$$

where x_{kl}^{\star} is the decision variable corresponding to A^{\star} , defined as

$$\mathbf{x}_{kl}^{\star} = \begin{cases} 0, (k, l) \notin A^{\star}, \\ 1, (k, l) \in A^{\star}, \end{cases}$$

r is the index of the haplotype containing reference. Thus, A^{\star} is a feasible solution of problem (8).

Second, we will show that "the output A^* of Algorithm 1 is an optimal solution of problem (8)". Let A be any feasible solution of problem (8). Then, by the constraints of problem (8), A can be represented as

$$A = \{(s(\rho(i)), s(i)) | i = 0, 1, \dots, m - 2\},$$
(13)

where $s(\rho(i))$ is the unique ancestor of s(i), for any i = 0, 1, ..., m - 2. Considering Equations (12) and (13), and Lemma 2, we have

$$w_2(e_{s(\pi^{\star}(i)),s(i)}) \prec w_2(e_{s(\rho(i)),s(i)}), \forall i = 0, 1, ..., m-2.$$

By Lemma 3, we have

$$\sum_{j=0,1,\dots,m-2} w_2(e_{s(\pi^\star(j)),s(j)}) \prec \sum_{j=0,1,\dots,m-2} w_2(e_{s(\rho(j)),s(j)})$$

that is $w_2(A^*) \prec w_2(A)$. Thus A^* is an optimal solution of problem (8). Q.E.D.

By sorting the haplotypes in step 1 of Algorithm 1, McAN avoids the evaluation of the condition $M_j \subsetneq M_i$ for any *i* and *j* satisfying $\pi^*(i) < j \le m-1$, and avoids storing the edges set *E* and the weight, including d_{ij} and \tilde{t}_{ij} , of all edges in *E*. The complexity of McAN depend on the selection of the sorting algorithms. If the quicksort is used in Algorithm 1, the best-case time complexity is reduced from $O(|V|^2)$ to $O(|V| \log |V|)$ and the space complexity is reduced from O(|E|) to O(|V|) in Algorithm 1, compared with Algorithm S1.

Compared with the other haplotype network construction algorithms, the worst-case time complexity of McAN is $O(m^2)$, which is equal to the worst-case time complexity of MSN and smaller than the time complexities of MJN ($O(n^{2.2})$) (average-case) [3] and TCS ($O(m^5)$) (worst-case) [26], where mis the number of haplotypes and n is the number of sequences.

RESULTS Performance testing on simulated datasets

We evaluated the accuracy of McAN by comparison with four existing popular algorithms MSN, MJN, TCS and RMST on a simulated dataset with 1001 sequences. We found that the AUC (area

Table 1. Evaluation of accuracy and memory cost on a simulateddataset including 1001 sequences

Algorithms	AUC	Memory cost (KB)
McAN	0.997	5844
MSN	0.998	59 960
MJN	0.997	63 676
TCS	0.997	62 928
RMST	0.998	114 725

under the curve) of McAN, MSN, MJN, TCS, and RMST are 0.997, 0.998, 0.997, 0.997 and 0.998, respectively, showing that McAN is as good as MJN and TCS, and is slightly lower than MSN and RMST (Supplementary Figure S2A-E, Table 1). We also evaluated the memory cost, and found that McAN consumed less than a tenth of the memory of the other four algorithms. On a large-scaled dataset including 20 001 sequences, the AUC of McAN remains 0.997 (Supplementary Figure S2F).

Performance testing on real SARS-CoV-2 datasets

The running performance of McAN was tested using 1 124 837 SARS-CoV-2 genome sequences retrieved from RCoV19 [7, 8] as of 26 July 2021 on a personal laptop (Intel Core i5-6200 U CPU and 8GB memory running with Ubuntu 20.04 operating system). We first established a small dataset by randomly sampling SARS-CoV-2 sequences. When the number of sequences ranges from 200 to 1000, McAN consistently obtained higher efficiency, running about 1000 times faster than, TCS and MJN and 100 times faster than RMST and MSN (Figure 2A). While on a larger dataset with different sequence counts ranging from 100 thousand to one million, even when sequence count reaches 1 million, McAN required less than 20 minutes, demonstrating that McAN is able to construct a haplotype network using large-scale dataset.

Given that tens of millions of SARS-CoV-2 sequences are currently available, the performance of paralleled McAN was tested on a server (2 Hygon C86 7185 32-core Processor, 512GB memory with CentOS Linux release 7.4.1708 (Core) operating system) using 4 990 399 SARS-CoV-2 sequences from RCoV19 as of 20 April 2022. When the number of threads ranges from 1 to 50, the running time of paralleled McAN rapidly declined from 28 696.5 to 1511.9 s (Figure 2B).

Performance evaluation using independent datasets

Independent datasets are usually used to assess the performance of a newly developed algorithm. Thus, we established two independent datasets for SARS-CoV-2 [9, 27] and evaluated performance of McAN. First, we tested on a real dataset of 482 SARS-CoV-2 genome sequences including 72 isolated from the Diamond Princess cruise and 410 from multiple countries/regions globally as detailed in [9]. Unlike MSN, TCS and MJN algorithms, McAN produced a direct evolutionary route from the reference (MN908947.3) to the Diamond Princess cluster (Supplementary Figure S3). This result is in agreement with a finding that SARS-CoV-2 dissemination on the Diamond Princess cruise is originated from a single introduction event [9]. Moreover, the haplotype network constructed by McAN using 130 major haplotypes of sublineages of L and S lineages from 121 618 SARS-CoV-2 genomes [27] shows a distinct delineation among these sublineages (Supplementary Figure S4), suggesting that McAN is capable of accurately tracking



Figure 2. Running time for MJN, TCS, MSN, RMST and McAN. (A) Comparison of running time for MJN, TCS, MSN, RMST and McAN with the number of sequences ranging from 200 to 1000 (single thread). (B) Running time of McAN with number of threads ranging from 1 to 50 for 4 990 399 sequences.

the evolution of SARS-CoV-2 during the development of the global pandemic of COVID-19.

In addition to SARS-CoV-2, we further tested McAN on other kinds of viruses including Monkeypox virus (MPXV) [28] and human influenza A viruses [29]. The haplotype network of monkeypox shows that most monkeypox samples collected in 2022 form a separate outbreak cluster except for two monkeypox samples collected from the USA in 2022 (Supplementary Figure S5A), supporting a previous study that 2022_FL001 and 2022_VA001 are unrelated to other cases occurring in 2022 in the USA. While using human influenza A viruses in 1918, we found that the main haplotype of these viruses consists of three samples collected in Germany and five samples collected in the USA (Supplementary Figure S5B). This finding strongly suggests that no geographic segregation exists between Europe and North America.

Web server guide and tools for haplotype network construction

To enable scholars to use McAN regardless of their computational skills, we established a free and user-friendly web server, which is available at https://ngdc.cncb.ac.cn/bit/hapnet/. On the webserver page, after a short introduction of the functions incorporated in McAN (Supplementary Figure S6), users can upload the genomic variants data either in VCF or customized genovar format, together with the corresponding metadata (Figure 3A). If the genomic data are from SARS-CoV-2, users can easily combine their data with a particular subset of SARS-CoV-2 sequences and metadata in RCoV19 by setting sampling date or country, or by sampling randomly. Next, users can filter sites by setting a minimum mutation rate. Haplotype network of user provided sequences will be constructed and community lineages will be determined meanwhile by hierarchical agglomeration algorithm [30] via McAN. The haplotype network results can be downloaded directly or viewed interactively in a viral haplotype network viewer developed in-house (Figure 3B), or sent via email.

Because of the bottleneck of network transmission, webservers cannot work when the number of inputted sequences is too large. To allow users to run McAN locally, we also developed an offline tool in C/C++ that can be freely downloaded from https:// ngdc.cncb.ac.cn/biocode/tools/BT007301 and https://github.com/ Theory-Lun/McAN. This tool supports input files in VCF format as well as customized genovar formats, and generate output files in TSV, JSON and GraphML formats, all of which are suitable for visualization and publication.

Interactive web-based visualization platform for haplotype network

We further developed an interactive web-based haplotype network visualization platform to facilitate tracing the genealogy, i.e. viral haplotype networks integrating spatial-temporal information. The platform features four major interconnected panels, each presenting spatial information, haplotype network, timeline and concise meta information of sequences. In spatial panel, it depicts colored circles that correspond to different locations, mirroring the colors in the haplotype network graph. The size of each circle reflects the number of samples in the current location, relative to the overall sample size. While on the network, it can be zoomed in and out and can be moved as a whole. Hovering over a node provides cluster information, while dragging a node makes it easier to observe and format. Clicking on a node highlights both the node and its branch for focused analysis, and the map and table dynamically update when nodes are clicked. For the timeline, the visibility of nodes can also be adjusted by dragging the timeline. With the 'PLAY' button, users can visualize how their sequences evolve over time in various locations. (Figure 3C). Additionally, lineages of evolving clusters are analyzed simultaneously and colored in different colors, which can be viewed by clicking on the 'View group' button (Figure 3D). All haplotype network figures can be saved in PNG format. This platform is capable of displaying networks with tens of thousands of nodes.

CONCLUSION AND DISCUSSION

This paper proposed McAN, a novel haplotype network construction algorithm and platform based on minimum-cost arborescence. McAN is capable for analyzing a large number of sequences and helpful for molecular tracing of pathogens for pandemics (e.g. COVID-19). McAN outperforms existing algorithms and achieves higher accuracy and efficiency in haplotype network construction by testing on multiple datasets. Its visualization platform enables the display of a haplotype network with tens of thousands of nodes across space and time, which offers multiple panels that present different facets of the network and remain synchronized while interacting. These results strongly suggest that McAN is a desirable platform for



Figure 3. Web server for haplotype network construction. (A) Homepage of McAN's web server, where users can upload their genome mutation data and metadata, and set parameters to construct haplotype network online. (B) In-house developed haplotype network viewer, allowing users to interactively view the haplotype network constructed by McAN. (C) In the haplotype network viewer, nodes of interested lineages can be highlighted. (D) Sequential screenshots showing interactive functions, where network nodes can be hidden or make visible by dragging the timeline bar below the viewer page.

analyzing massive dynamic data, especially SARS-CoV-2 datasets, and will be beneficial to the study of genome evolution.

While McAN is clearly useful for constructing haplotype network of viruses, it is limited in a number of ways. First, the mutation spectrum history (mutations in ancestry haplotype should be contained in descendant haplotype) implies that McAN cannot detect any reverse mutations, which have been occurred in SARS-CoV-2 [31]. Second, we used arborescence to represent a haplotype network and therefore McAN cannot detect any recombination, which have been reported in the Omicron variant of Covid-19 virus [32, 33]. Third, the preprocessing of sequences and selection of root might affect the edges of the haplotype network constructed by McAN. User should use a high-quality sequence as the reference. If the quality of reference is low; i.e. the length of sequence is short or the number of ambiguous nucleotides is large, the accuracy of mutation detection may decrease. This will further affect the accuracy of haplotype network. In addition, we advise choosing the earliest sequence as reference. If a relatively late sequence is used as reference, the direction of edges in haplotype network may not reflect the direction of evolution correctly.

Key Points

- Minimum cost arborescence network (McAN) algorithm constructs haplotype network by considering mutation spectrum history, node size and sampling time simultaneously.
- McAN platform provides a user-friendly web server as well as an offline tool, allowing users to construct haplotype network, detect lineages and visualize the results interactively online.
- McAN is capable to build haplotype network for millions of sequences data with high accuracy and efficiency (about 20 minutes for millions of sequences using one to fifty threads).
- McAN is helpful for the molecular tracing of pathogens for pandemics and is beneficial to deepening the understanding of viruses including but not limited to SARS-CoV-2, Monkeypox virus and human influenza A viruses.

SUPPLEMENTARY DATA

Supplementary data are available online at https://academic.oup. com/bib.

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